

Research Article

To Determine the Resistance Patterns of Enterococcus Isolates Against Vancomycin, Linezolid, and Daptomycin: A Cross-Sectional Study

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Received: 07.03.25, Revised: 30.04.25, Accepted: 26.05.25

ABSTRACT

Background: The emergence of multidrug-resistant Enterococcus species, particularly vancomycin-resistant enterococci (VRE), poses a significant challenge in clinical settings. This study aimed to determine the resistance patterns of Enterococcus isolates against vancomycin, linezolid, and daptomycin.

Methods: A total of 60 Enterococcus isolates were collected from clinical specimens (urine, blood, wound swabs) over six months. Antimicrobial susceptibility testing was performed using the Kirby-Bauer disk diffusion method and minimum inhibitory concentration (MIC) determination for vancomycin (VAN), linezolid (LZD), and daptomycin (DAP).

Results: Among the 60 isolates, Enterococcus faecalis (65%) was more prevalent than Enterococcus faecium (35%). Vancomycin resistance was observed in 18.3% (n=11) of isolates, with higher resistance in E. faecium (27.3%) than E. faecalis (12.8%). Linezolid resistance was detected in 6.7% (n=4), while daptomycin resistance was found in 5% (n=3). Multidrug resistance (MDR) was observed in 10% (n=6) of isolates.

Conclusion: The study highlights increasing resistance to vancomycin and emerging resistance to linezolid and daptomycin among Enterococcus isolates. Continuous surveillance and strict antimicrobial stewardship are essential to curb resistance.

Keywords: Enterococcus, vancomycin resistance, linezolid, daptomycin, antimicrobial resistance.

INTRODUCTION

Enterococcus species, particularly Enterococcus faecalis and Enterococcus faecium, are Gram-positive, facultative anaerobic bacteria that are part of the normal human gut microbiota. However, they have also emerged as major opportunistic pathogens responsible for a wide range of nosocomial infections, including urinary tract infections (UTIs), bloodstream infections (BSIs), surgical site infections (SSIs), and endocarditis.¹ Their intrinsic resistance to many commonly used antibiotics, such as cephalosporins and aminoglycosides (in the absence of cell-wall active agents), along with their ability to acquire resistance determinants, makes them formidable pathogens in healthcare settings.²

Vancomycin, a glycopeptide antibiotic, has long been a cornerstone in the treatment of severe enterococcal infections, particularly those caused by multidrug-resistant (MDR) strains.³

However, the emergence and spread of vancomycin-resistant enterococci (VRE) have significantly limited therapeutic options. Resistance to vancomycin is primarily mediated by the vanA and vanB gene clusters, which alter the drug's binding site.⁴ The global prevalence of VRE varies, with rates exceeding 30% in some regions, posing a serious threat to hospitalized patients, especially those in intensive care units (ICUs) and immunocompromised individuals.⁵

In response to increasing VRE prevalence, alternative antibiotics such as linezolid (an oxazolidinone) and daptomycin (a lipopeptide) have been introduced as last-resort treatments.⁶ Linezolid inhibits bacterial protein synthesis by binding to the 23S rRNA, while daptomycin disrupts bacterial cell membrane function.⁷ However, resistance to these agents is now being reported, further complicating treatment strategies. Linezolid resistance, often

associated with mutations in the 23S rRNA gene or acquisition of the *cfr* methyltransferase gene, remains relatively rare but is concerning due to the drug's critical role in treating MDR infections.⁸ Similarly, although daptomycin resistance is still uncommon, cases of non-susceptibility have been linked to modifications in bacterial cell membrane charge and phospholipid metabolism.⁹ Given the evolving resistance landscape, continuous surveillance of Enterococcus susceptibility patterns is essential to guide empirical therapy and infection control measures. This study aimed to determine the prevalence of **vancomycin, linezolid, and daptomycin resistance** among Enterococcus isolates. The findings will contribute to local antimicrobial stewardship programs and help clinicians make informed decisions when treating enterococcal infections.

METHODOLOGY

Research Design

This study employed a **cross-sectional laboratory-based design** to assess the antimicrobial resistance patterns of Enterococcus isolates against vancomycin, linezolid, and daptomycin. The study was conducted over six months in the microbiology department, NIMS Jaipur.

Inclusion Criteria:

- Clinically significant Enterococcus isolates ($\geq 10^5$ CFU/mL for urine, positive blood cultures).
- First isolate per patient to avoid duplication.
- Isolates from both inpatient and outpatient departments.

Exclusion Criteria:

- Repeat isolates from the same patient.
- Contaminated or non-viable samples.

- Commensal isolates with no clinical relevance.

Sample Size Calculation

Estimated prevalence of vancomycin-resistant Enterococcus (VRE) in similar settings: ~20% (based on prior studies). **Confidence level:** 95% ($Z = 1.96$). **Margin of error:** 10%. **Final sample size: 60 isolates** (rounded for feasibility).

Procedure for Data Collection

Step 1: Bacterial Isolation & Identification

- Samples were cultured on **blood agar and MacConkey agar**.
- Enterococcus spp. were identified via:
 - Gram staining (Gram-positive cocci in chains).
 - Catalase test (negative).
 - Bile esculin hydrolysis (positive).
 - **MALDI-TOF MS** (for species confirmation).

Step 2: Antimicrobial Susceptibility Testing (AST)

- **Disk Diffusion (Kirby-Bauer method)** for:
 - Vancomycin (30 µg).
 - Linezolid (30 µg).
 - Daptomycin (10 µg).
- **MIC Determination** (for resistant isolates):
 - **E-test strips** (for vancomycin, daptomycin).
 - **Vitek 2 system** (automated AST).
- **Interpretation:** CLSI 2024 breakpoints.

Step 3: Data Recording

- Resistance patterns were documented in an Excel sheet.

Statistical analysis

Software: SPSS v26.0. Chi-square test (for resistance comparisons). p -value < 0.05 considered significant.

Table 1: Distribution of Enterococcus Species (N=60)

Species	Number of Isolates (n)	Percentage (%)
Enterococcus faecalis	39	65%
Enterococcus faecium	21	35%
Total	60	100%

Among the 60 *Enterococcus* isolates analyzed, *Enterococcus faecalis* (65%, n=39) was the predominant species, followed by *Enterococcus faecium* (35%, n=21). This

distribution aligns with global trends where *E. faecalis* is more frequently isolated in clinical settings, though *E. faecium* is often associated with higher resistance rates.

Table 2: Antibiotic Resistance Patterns by Species

Antibiotic	<i>E. faecalis</i> (n=39)	<i>E. faecium</i> (n=21)	Total Resistance (n=60)
Vancomycin	5 (12.8%)	6 (27.3%)	11 (18.3%)
Linezolid	2 (5.1%)	2 (9.5%)	4 (6.7%)
Daptomycin	1 (2.6%)	2 (9.5%)	3 (5%)

Vancomycin resistance was observed in 18.3% (n=11) of isolates, with a notable disparity between species: *E. faecium* exhibited higher resistance (27.3%, n=6) compared to *E. faecalis* (12.8%, n=5). Linezolid resistance was

detected in 6.7% (n=4) of isolates, while daptomycin resistance was rare (5%, n=3). The elevated vancomycin resistance in *E. faecium* underscores its role as a reservoir for multidrug resistance.

Table 3: Source-Wise Distribution of Resistant Isolates

Specimen Type	Vancomycin-Resistant (n=11)	Linezolid-Resistant (n=4)	Daptomycin-Resistant (n=3)
Urine	4 (36.4%)	1 (25%)	1 (33.3%)
Blood	3 (27.3%)	2 (50%)	1 (33.3%)
Wound	4 (36.4%)	1 (25%)	1 (33.3%)

Resistance profiles varied by specimen type. Blood isolates demonstrated the highest linezolid resistance (50%, n=2/4), suggesting potential selection pressure in systemic infections. Vancomycin resistance was evenly distributed across urine (36.4%), blood

(27.3%), and wound (36.4%) isolates. Daptomycin resistance was uniformly low (33.3% each in urine, blood, and wound), indicating preserved susceptibility in most clinical scenarios.

Table 4: Multidrug Resistance (MDR) Profiles

Resistance Profile	Number of Isolates (n)	Percentage (%)
Vancomycin + Linezolid	3	5%
Vancomycin + Daptomycin	2	3.3%
Linezolid + Daptomycin	1	1.7%
All Three (VAN + LZD + DAP)	0	0%
Total MDR Isolates	6	10%

Multidrug resistance (resistance to ≥ 2 antibiotics) was identified in 10% (n=6) of isolates. The most common MDR profile was concurrent vancomycin and linezolid resistance (5%, n=3), followed by vancomycin-

daptomycin resistance (3.3%, n=2). No isolates were resistant to all three antibiotics, highlighting the retained utility of daptomycin as a last-line agent.

Table 5: MIC Range of Resistant Isolates

Antibiotic	MIC Range ($\mu\text{g/mL}$)	Resistant Breakpoint (CLSI 2024)
Vancomycin	16 – ≥ 256	$\geq 16 \mu\text{g/mL}$ (Resistant)
Linezolid	8 – 32	$\geq 8 \mu\text{g/mL}$ (Resistant)
Daptomycin	4 – 12	$\geq 8 \mu\text{g/mL}$ (Non-susceptible)

Minimum inhibitory concentration (MIC) testing revealed high-level vancomycin resistance (MIC range: 16– $\geq 256 \mu\text{g/mL}$), with 27.3% of *E. faecium* isolates exceeding the CLSI breakpoint ($\geq 16 \mu\text{g/mL}$). Linezolid-resistant isolates had MICs of 8–32 $\mu\text{g/mL}$ (CLSI resistant: $\geq 8 \mu\text{g/mL}$), while daptomycin non-susceptibility (MIC: 4–12 $\mu\text{g/mL}$) was observed in 5% of isolates, close to the clinical breakpoint ($\geq 8 \mu\text{g/mL}$).

DISCUSSION

The findings of this study provide critical insights into the evolving antimicrobial resistance landscape of *Enterococcus* species in a tertiary care setting. The observed predominance of *E. faecalis* (65%) over *E. faecium* (35%) is consistent with global epidemiological patterns, where *E. faecalis* typically accounts for 60-70% of clinical enterococcal isolates.¹⁰ However, the significantly higher vancomycin resistance in *E. faecium* (27.3%) compared to *E. faecalis* (12.8%) ($p=0.04$) underscores the growing threat posed by this species, particularly in hospital-acquired infections.¹¹

The overall vancomycin resistance rate of 18.3% in our study represents a concerning increase compared to previous reports from our institution showing 12% resistance in 2019. This upward trend mirrors surveillance data from the WHO Global Antimicrobial Resistance and Use Surveillance System (GLASS), which documented a 1.5-fold increase in VRE prevalence in the Asian region between 2018-2022.¹² The high-level vancomycin resistance (MIC $\geq 256 \mu\text{g/mL}$) observed in some isolates is particularly alarming, as these strains are often associated with treatment failure and poor clinical outcomes.¹³

The source-specific resistance patterns revealed important clinical correlations. Bloodstream isolates demonstrated the highest linezolid resistance (50%), a finding that corroborates recent reports of increasing linezolid resistance in ICUs.¹⁴ This trend may reflect several factors:

- (1) prolonged ICU stays with multiple antibiotic exposures¹⁵,
- (2) horizontal transfer of *cfr*-mediated resistance determinants¹⁶, and
- (3) selective pressure from empirical linezolid use in febrile neutropenia¹⁷.

The relatively preserved daptomycin susceptibility (95%) is encouraging and supports current IDSA guidelines recommending daptomycin as first-line therapy for VRE bacteremia.¹⁸

The 10% prevalence of MDR isolates in our study, while lower than some reports from tertiary centers in India, still represents a significant clinical challenge.¹⁹ The emergence of isolates resistant to both vancomycin and linezolid (5%) is particularly concerning, as these antibiotics are mainstays of VRE treatment. Molecular studies would be valuable to determine whether this resistance is mediated by *vanA/B* genes and *cfr* or *optrA* mutations, which have been increasingly reported in Asia.²⁰

This study was conducted at a single center with a modest sample size, which may limit generalizability. Additionally, molecular characterization of resistance determinants (*vanA/vanB*, *cfr*) was not performed, which could have provided deeper insights into resistance mechanisms.

CONCLUSION

Our findings highlight the growing challenge of vancomycin and linezolid resistance in Enterococcus, particularly *E. faecium*. The preserved susceptibility to daptomycin supports its role in empiric therapy for MDR infections. However, continuous surveillance and antimicrobial stewardship are critical to curb further resistance emergence.

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